

STATE OF CONNECTICUT DEPARTMENT OF ENERGY AND ENVIRONMENTAL PROTECTION

Robert Klee Commissioner

Final

State Wildlife Grants F14AF01186 (T-15-R-1) Interim Performance Report September 1, 2014 – June 30, 2018

Restoration of Anadromous Alewife to Lakes of Connecticut

State Wildlife Grant F14AF01186 (T-15-R-1) Restoration of Anadromous Alewife to Lakes of Connecticut

Final Performance Report

Principal Investigators:

David M Post, Lead, Yale University

Eric P. Palkovacs, University of California – Santa Cruz

Project Goal: To determine the impact of landlocked alewives on anadromous alewife restoration in Rogers Lake, Old Lyme, CT.

In the spring of 2014, the CT DEEP opened the final of three fishway on Mill Brook allowing anadromous alewife (river herring) access to historic spawning grounds in Rogers Lake. In Rogers Lake there was already a resident landlocked population. A decade of research on alewife in Rogers Lake and other lakes in the region has shown that anadromous and landlocked alewife differ in traits that strongly affect the ecology of lakes, including the duration in freshwater, gape width, gillraker spacing, prey size-selectivity, diet composition, habitat usage, and whole-body morphology (Palkovacs et al. 2008, Palkovacs and Post 2008, Schielke et al. 2011, Jones et al. 2013, Palkovacs et al. 2014). The fishway allowed for anadromous and landlocked alewife to come into "secondary contact," a process in which previously isolated and potentially divergent linages come back into contact. At the start of this project, the outcome of secondary contact between landlocked and anadromous alewife was uncertain. Landlocked alewife could outcompete and prevent establishment of anadromous alewife, anadromous alewife could establish a population that coexists with the landlocked population, or landlocked and anadromous alewife could hybridize in the lake. These potential outcomes would have important implications for anadromous alewife restoration efforts in CT and across the range of anadromous alewife. For example, a large locally adapted resident landlocked population might outcompete a small colonizing anadromous population, in which case additional management activities would be needed to facilitate anadromous alewife restoration. The outcomes of restoration and secondary contact could also have implications for the food webs and water quality of lake with existing landlocked populations and restored anadromous populations.

This project was designed to track the ecological and evolutionary implication of restoring spawning access for anadromous alewife in Rogers Lake, CT. This research tracked the number and genotypes of spawning anadromous adults entering the lake. It evaluated the potential for hybridization by analyzing spawning time overlap between anadromous and landlocked alewife populations in the region. Once anadromous alewife were in Rogers Lake, it used genetic data to assign young-of-the-year fish as anadromous, landlocked, or hybrids. These classifications

allowed the evaluation of spawning success and juvenile production, key metrics for river herring recovery.

The research funded by this CT DEEP SWG grant provided the critical proof of concept and preliminary data that the PIs utilized to obtain funding from the National Science Foundation (NSF) to continue this and related research through at least 2020.

Major accomplishments:

- (1) We tracked the number and genotypes of all anadromous alewife entering Rogers Lake in 2014 2018.
- (2) We combined juvenile density estimates with genetic assignments to determine the abundance of anadromous alewives, landlocked alewives, and anadromous-landlocked hybrids in Rogers Lake following restoration.
- (3) We use otolith-based aging to determine spawning time for anadromous, landlocked, and hybrid alewives to assess the effects of spawning time overlap on potential rates of hybridization.
- (4) We evaluate the effects of restoration on body size and habitat use by landlocked, anadromous, and hybrid young-of-the-year alewife.
- (5) We started to evaluate the effects of restoration on the Rogers Lake food web and water quality (a core long-term question for our NSF funded research).

Number, phenotype, and genotypes of adult anadromous alewife entering Rogers Lake:

The fishway into Rogers Lake was opened in the spring of 2014. Our initial goal was to allow anadromous alewife to naturally re-colonize Rogers Lake. Unfortunately, the anadromous alewife run below the fishway declined to such low abundances in 2014 that no fish successfully passed the fishway into Rogers Lake. As our backup plan, in 2015, in collaboration with the CT DEEP we started stocking adult anadromous alewife into Rogers Lake (134 in 2015; 1144 in 2016; 2787 in 2017; 3392 in 2018). In 2016-2018 we stocked fish from nearby Bride Lake, and stocking was conducted on three dates in 2017 and 2018 to capture the range of genotypes and represent the range of variation in the timing of spawning. All fish were measured for length, sexed, and fin clipped for genetic analysis. One major advantage of stocking was that it provided us the ability to measure and genotype every single adult anadromous alewife that entered Rogers Lake.

In 2018, we saw signs that the anadromous alewife run downstream of the Rogers fishway had started to recover (6,900 fish passed a CT DEEP monitored fishway downstream of the Rogers Lake fishway) but none of those fish passed through the intervening spawning habitat to ascend into Rogers Lake in 2018. We will continue to stock adult alewife into Rogers Lake and with our CT DEEP colleagues continue to monitor upstream passage of adult alewife as part of our ongoing NSF funded research.

We developed a novel set of 'microhaplotype' genetic markers for specific use in this project. Microhaplotypes are short DNA fragments containing multiple single nucleotide polymorphisms

(SNPs). Each microhaplotype contains 2 – 5 SNPs per fragment, which in turn leads to more information per locus. The major technical advantages of this technique are more information per locus and the ability to account for any new variants detected. Power analysis confirmed the reliability of these markers to distinguish landlocked, anadromous, and hybrid individuals with very high confidence.

Since 2015, we have measured sex and length on all of the nearly 7,500 adult anadromous alewife entering Rogers Lake, and genotyped all of the fish entering the lake between 2015 and 2017 (Table 1). These data were used to assign juveniles as landlocked, anadromous, or hybrids (see below), and will allow us to ultimately conduct parentage analysis on juvenile fish and evaluate spawning success based on sex, length, and timing of stocking (e.g., did late stocked anadromous alewife contribute more to the production of hybrids than early stocked anadromous alewife).

Table 1. Number of adult anadromous alewife stocked into Rogers Lake, sex ratio, mean length and status of genetic analysis.

Year	Adults Stocked	Sex ratio (m:f)	Mean length (mm)	Extracted	Genotyped
2015	134	2.62	281	134	134
2016	1144	0.60	272	1144	1144
2017	2787	1.39	264	2787	2787
2018 [†]	3392	1.22	268	49	

[†] The 2018 adult alewife will be extracted and genotyped in the fall of 2018 as part of the ongoing NSF funded research.

Spawning time overlap:

Overlap in spawning time is one main factor that may limit the opportunity for hybridization between landlocked and anadromous alewife populations. We used date on anadromous alewife run time collected since the early 2000s, and spawning date estimates for anadromous and landlocked alewife populations in 2013-2015 (otolith back-calculated ages and temperature-dependent estimates of development time within the egg) to estimate the potential for spawning overlap between anadromous and landlocked alewife populations. We found that anadromous alewife spawned earlier and over a shorter duration than landlocked alewife. In 2014 and 2015, the mean spawn date for anadromous alewives occurred on May 19, whereas mean spawn data for landlocked alewives occurred on June 29. For all populations and in all years, our results estimate that 3% - 13% of landlocked alewife spawned within a period of overlap with anadromous alewife. The degree of spawning time overlap was primarily driven by yearly variation in landlocked alewife spawning time (and specifically the initiation of spawning), whereas the timing and duration of spawning for anadromous alewife runs was found to be stable across years. Our results on spawning time overlap were published in Littrell et al. (2018) (doi:10.1111/eva.12645), which is attached to this final report. For Rogers Lake

specifically, we estimate around 3% overlap in the distributions of spawning time, indicating the potential for limited hybridization between landlocked and anadromous alewife re-colonizing Rogers Lake. As we note in Littrell et al. (2018), there is considerable interannual variation within a lake and variation among lakes in the date of initiation of spawning for landlocked alewife and this produces a range of potential overlap that could create interannual and among lake variation in hybridization rates after secondary contact.

Juvenile genotypes:

Starting in 2015, we collected juvenile alewife in Rogers Lake and our references lakes to evaluate spawning success using our genetic markers, and to estimate densities and habitat use. Most of the juveniles are young-of-the-year but, as we note below, some appear to be 1+ individuals that have held over winter in Rogers Lake.

We collect juvenile alewife in August of each year using replicate sets of a research purse sein designed to encircling 100m^2 . We sample at night because alewife move up in the water column and spread out across the lake at night, allowing us to more effectively estimate population densities. Previous research documented different patterns of habitat use for anadromous and landlocked juveniles (Schielke et al. 2011, Jones et al. 2013). Landlocked alewife typically have slightly higher densities offshore than inshore and diets are composed of nearly 100% pelagic prey. In contrast, anadromous alewife, particularly at higher densities, move inshore, where they can reach very high densities compared to off shore habitat, and can have diets that range from nearly 100% pelagic to nearly 100% littoral prey. Because of these differences, we sampled both inshore and offshore habitats.

We collected and genotyped nearly 3,000 juvenile alewife from Rogers Lake from 2015-2017. In 2015, we found only landlocked alewife. In 2016, we found a small number of juvenile anadromous alewife in Rogers Lake (Table 2), indicating the first successful reproduction of anadromous alewife in Rogers Lake in recent history. In 2017, we found a larger proportion of anadromous alewife and for the first time anadromous-landlocked hybrid juveniles in Rogers Lake (Table 2). About half of the hybrid individuals in 2017 were from anadromous adults

Table 2. Number of juvenile alewife collected in Rogers Lake, status of genetic analysis, and genetic identification.

	Juveniles			Percent†		
Year	collected	Extracted	Genotyped	Landlocked	Anadromous	Hybrid
2015	450	450	450	100%	0%	0%
2016	1120	1120	1120	99.6%	0.4%	0%
2017	1420	1420	1420	89.1%	6.5%	4.4%

[†] Percentage of juveniles genotypes successfully. We had a 2.7% rate of genotyping failure due to degraded DNA in 2017. These percentages are different than those for the total population size below which are slightly different because they are based on inshore and offshore distributions.

stocked in 2017, and half were backcrosses. Backcrosses have one pure anadromous or landlocked parent and one hybrid parent, likely from 2016. This means there must have been hybrids produced in 2015 or 2016 (we believe 2016 is most likely) and those hybrids both remained in Rogers Lake over winter and were mostly likely reproductively active as 1+ individual (spawning at 1+ is not uncommon for landlocked alewife). We are still in the process of conducting the parentage analysis (it is quite complex) and we expect to be able to identify all of the anadromous parents or grandparents of the backcrosses, which will tell us in which year the hybrid parents were produced.

The low rate of hybridization that we observed in Rogers Lake in 2017 is consistent with the estimated percent of overlap (around 3%) predicted by our spawning time analysis (Littrell et al. 2018).

Juvenile population size:

We combine juvenile density estimates with genetic assignments to determine the population size of anadromous, landlocked alewives, and hybrids in Rogers Lake in 2013-2017. We found considerable interannual variation in juvenile alewife population size (Figure 1, Table 3). 2015 and 2017 were years of very high juvenile abundance, while 2016 was a year of small juvenile population size. In 2016, Rogers Lake produced around 90,000 landlocked alewife and, for the first time in recent history, around 6,500 anadromous alewife. As we noted above, there must have been hybrids produced in either 2015 or most likely in 2016. If they were produced in 2016, based on our catch rates for anadromous alewife, there were likely fewer than 2,000 hybrids in 2016. In 2017, Rogers Lake produced around 350,000 landlocked alewife, nearly 70,000 anadromous alewife, and nearly 23,000 landlocked-anadromous hybrid alewife.

Table 3. Population estimates for landlocked, anadromous, hybrid alewife in 2013 - 2017. Estimates are mean \pm 95% confidence intervals. Estimates are based on and weighted by inshore and offshore purse seine catch and genotype assignment.

Year	Landlocked	Anadromous	Hybrid
2013	223,483	0	0
	(122,412 - 324,555)		
2014	127,642	0	0
	(68,382 - 186,901)		
2015	431,067	0	0
	(343,975 - 518,159)		
2016	92,486	6,448	0
	(41,888 - 143,083)	(2,166 - 10,730)	
2017	350,186	68,527	22,953
	(172,561 - 527,812)	(44,711 - 92,343)	(12,741 - 33,165)

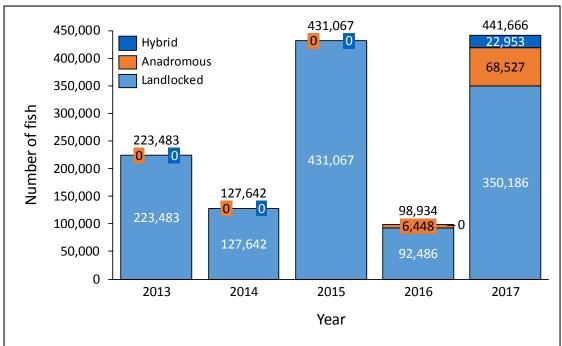


Figure 1. Estimated number of landlocked, anadromous and hybrid alewife in Rogers Lakes 2013 - 2017.

Juvenile habitat use:

In 2016 and 2017, most of the anadromous alewife were captured inshore, landlocked were captured predominately offshore, and, in 2017, hybrids were nearly evenly distributed among inshore and offshore habitats (Table 4). This is consistent with the general patterns of habitat use that we find for isolated landlocked and anadromous populations (Schielke et al. 2011, Jones et al. 2013). It appears that anadromous alewife are better able to utilize littoral (nearshore) resources than landlocked alewife. As part of our ongoing NSF funded research, we will be analyzing diet and resource use (using stable isotopes) of the landlocked, anadromous, and hybrid juveniles from 2016, 2017 and 2018.

Table 4. Proportion of the 2016 and 2017 inshore and offshore catch assigned to landlocked, anadromous, or hybrid genotypes.						
Year	ar Habitat Landlocked Anadromous Hybrid					
2016	Inshore	87.5%	12.5%	0%		
Offshore 99.9% 0.1% 0%						
2017	Inshore	59.1%	34.1%	6.8%		
Offshore 90.1% 5.6% 4.3%						

Juvenile body size:

We found considerable differences in the length of juvenile landlocked alewife among years and, in 2017, differences in length among landlocked, anadromous, and hybrid alewife (all August data; Figure 2). The variation in landlocked length in 2013 - 2016 was caused by variation in spawning date (Littrell et al. 2018). Interestingly, that suggests landlocked alewife spawned earlier in 2017 than in previous year (see below for further discussion). This may reflect the greater year-to-year variation in the initiation date of spawning for landlocked alewife as observed by Littrell et al. (2018), or it is possible that the early spawning of anadromous alewife in 2017 caused landlocked alewife to initiate spawning sooner than in previous year. We should be able to answer this question once we analyze spawning date data for Rogers Lake in 2018 and 2019 as part of our ongoing NSF funded research.

In 2017, anadromous alewife were longer than landlocked alewife (Figure 2), reflecting some combination of earlier spawning time (as we found in Littrell et al. 2018) and potentially faster growth rates. As part of our ongoing NSF funded research, we are using otoliths to age these fish so we can determine the relative impact of differences in spawning date and growth rate on the differences in length observed in 2017 and 2018. In 2017, one hybrid and three landlocked alewife were nearly twice as long as all of the other alewife (Hybrid: 158mm; Landlocked: 141-144 mm) suggesting they were 1+ or older and likely reproductively active. A combination of otolith and parentage analyses will allow us to determine the year in which the hybrid individual hatched (2016 or 2017).

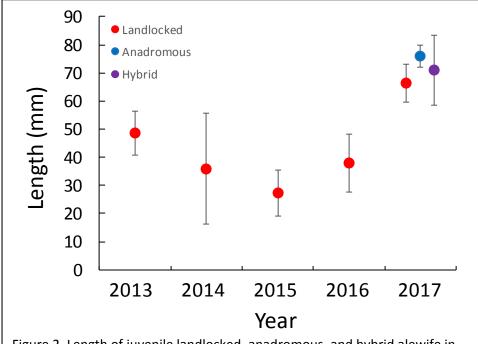


Figure 2. Length of juvenile landlocked, anadromous, and hybrid alewife in 2013 - 2017 (mean \pm standard deviation). There were too few anadromous and hybrid individuals in 2016 to calculate the mean length.

We can compare the length of Rogers Lake anadromous alewife in 2017 to the length of anadromous alewife in Bride Lake and Dodge Pond (all August data; Figure 3). Anadromous alewife in Rogers Lake were considerably larger than anadromous alewife in both Bride and Dodge (Figure 3). It is possible that the greater length of juvenile anadromous alewife in Rogers

Lake is a results of reduced competition with other anadromous alewife. Because anadromous alewife utilize inshore resources that are not utilized by landlocked alewife, they may be competing with each other more than with landlocked alewife. In 2017, the inshore density of anadromous alewife in Rogers Lake (0.1 fish/m²) was considerably lower than the inshore density of anadromous alewife in Bride (0.75 fish/m²) and Dodge (0.28 fish/m²), which may have allowed for greater growth rates in Rogers Lake in 2017.

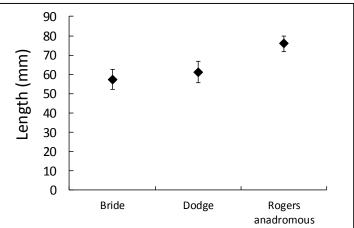


Figure 3. Length of Rogers Lake juvenile anadromous alewife compared to the length of Bride Lake and Dodge Pond anadromous alewife in August of 2017 (mean ± standard deviation).

We can also compare the length of landlocked alewife in Rogers Lake to the length of landlocked alewife in Pattagansett and Quonnipaug Lakes in 2013 - 2017 (all August data; Figure 4). We know from our spawning time analysis (Littrell et al. 2018) that landlocked alewife in Quonnipaug and Rogers lakes spawn at about the same time and grow at about the same

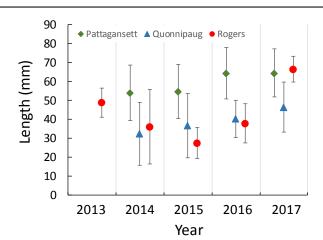


Figure 4. Length of juvenile landlocked alewife in Rogers Lake compared to juvenile landlocked alewife in Pattagansett and Quonnipaug lakes in 2013 - 2017 (mean ± standard deviation).

rate, and we see strong congruence in length between Quonnipaug and Rogers landlocked alewife in 2014 - 2016. We also know that Pattagansett Lake landlocked alewife typically spawn earlier than the Rogers Lake landlocked alewife and therefore reach greater length by August, as we see in our data for 2014 - 2016. In 2017, however, landlocked alewife from Rogers Lake are much larger than those found in Quonnipaug and were a similar size as those collected in Pattagansett. This supports our speculation above that the Rogers Lake landlocked alewife may have spawned earlier in 2017 than in the four previous years. We will be able to

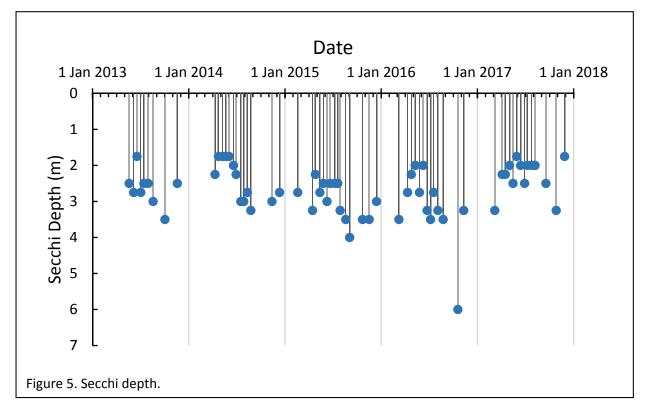
confirm this once we age the 2017 and 2018 alewife as part of our ongoing NSF funded research.

Preliminary impacts on the Rogers Lake food web and water quality

Landlocked and anadromous alewife differ in a number of ecologically relevant traits including the duration in freshwater, whole-body and feeding morphology, foraging behavior, habitat use, and migration patterns (Palkovacs and Post 2008, Post et al. 2008, Schielke et al. 2011, Jones et al. 2013). These differences have altered the ecological role of these two alewife forms such that landlocked and anadromous alewife have very different effects on community structure and ecosystem function (Post et al. 2008, Palkovacs and Post 2009, Walters et al. 2009, West et al. 2010, Twining et al. 2013, Weis and Post 2013). We do not know what to expect for the food web and ecosystem effects of the restoration of anadromous alewife to Rogers Lake, but from previous research we expect mean crustacean zooplankton length and algal biomass (measured as Chl. a concentrations, or potentially as Secchi depth) should be sensitive to changes in alewife dynamics.

Building upon 15 years of routine sampling, we collected zooplankton, measured Secchi depth, and estimated algal biomass using Chl. a concentrations every other week from April to September, and monthly (when possible) from October until March in 2013 - 2017.

<u>Water clarity</u> - We observed little if any change in Secchi depth in Rogers Lake in 2013-2017, with the exception of a remarkably clear date in October of 2016 (Figure 3).



<u>Algal biomass</u> - We measured little to no change in Chl. a concentrations in Rogers Lake in 2013 - 2017 (Figure 4). There has been no directional change in Chl. a concentrations from 2013 - 2017 (slope = 0.00; p = 0.40; $R^2 = 1.1$), although it appears that the mean growing season Chl. a concentration (June - September) was higher in 2017 than in previous years. Mean growing season Chl. a concentrations ranged from 3.75m in 2013 to 5.4m in 2018.

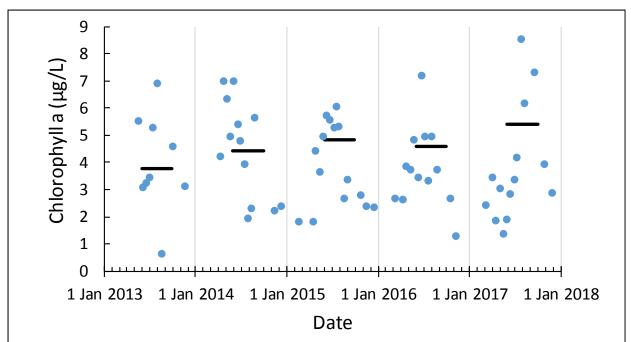


Figure 6. Chlorophyll a concentrations on each sampling date. The black line is the mean growing season (June - September) Chl. a concetrations for 2013 - 2017.

Zooplankton community - There have been changes in mean crustacean (Copepod and Cladoceran) zooplankton length in Rogers Lake (Figure 5). There was significant interannual differences in mean zooplankton length (one-way ANOVA; F_{4,56} = 9.84; p < 0.001) caused by 2017, which had a significantly greater mean zooplankton length than 2013 - 2016 (Bonferroni corrected post hoc tests at an overall alpha of 0.05). The summer of 2017 was the first year in our 15 years of sampling that we found a high densities of *Daphnia* spp. in Rogers Lake. *Daphnia* persisted in Rogers Lake until September. *Daphnia* are the largest bodied and most effective grazers in lakes, and are therefore important regulators of water quality and for the flow of energy to upper trophic levels. *Daphnia* were not present in high densities in 2018 so 2017 may have been an anomalous year. The high densities of *Daphnia* may explain the greater length of all genotypes of alewife in 2017.

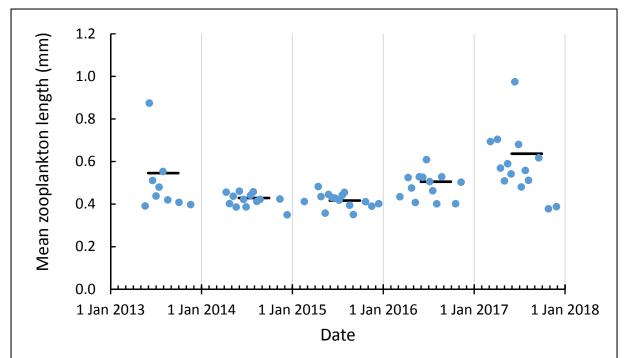


Figure 7. Mean crustacean zooplankton langth on each sampling date The black line is the mean growing season (June - September) crustacean zooplantkon length for 2013 - 2017.

Conclusion:

There are several key findings from our research on the restoration of Rogers Lake.

- 1. Anadromous alewife spawned successfully in Rogers Lake in both 2016 and 2017.
- 2. Anadromous and landlocked alewife produced a large number of hybrid offspring in 2017.
- 3. Some of the hybrid offspring collected in 2017 were backcrosses, which means first generation hybrids must have a) been produced in 2015 or 2016, b) remained in Rogers Lake over winter, and c) became reproductive in 2017 as 1+ or 2+ individuals.
- 4. Anadromous and hybrid alewife reached lengths that were similar to or larger than landlocked alewife in 2017, indicating that anadromous and hybrid alewife grow well even in a lake with considerable densities of locally adapted landlocked alewife.
- 5. Anadromous alewife were found at higher densities in inshore than offshore habitats. This is consistent with our observations in isolated anadromous populations, which heavily utilize inshore resource, particularly at higher densities. The utilization of inshore resources in Rogers Lake may reduce direct competition with the resident landlocked population in Rogers Lake.
- 6. There is little evidence for significant changes in the Rogers Lake food web. We will need additional years of sampling to determine if the increase in large-bodied zooplankton (*Daphnia*) observed in 2017 was anomalous or a systematic change in the zooplankton community.

References:

- Jones, A. W., E. P. Palkovacs, and D. M. Post. 2013. Recent parallel divergence in body shape and diet source of alewife life history forms. Evolutionary Ecology **27**:1175-1187.
- Littrell, K. A., D. Ellis, S. R. Gephard, A. D. MacDonald, E. P. Palkovacs, K. Scranton, and D. M. Post. 2018. Evaluating the potential for pre-zygotic isolation and hybridization between landlocked and anadromous alewife (Alosa pseudoharengus) following secondary contact. Evolutionary Applications.
- Palkovacs, E. P., K. B. Dion, D. M. Post, and A. Caccone. 2008. Independent evolutionary origins of landlocked alewife populations and rapid parallel evolution of phenotypic traits. Molecular Ecology **17**:582-597.
- Palkovacs, E. P., E. G. Mandeville, and D. M. Post. 2014. Contemporary trait change in a classic ecological experiment: rapid decrease in alewife gill-raker spacing following introduction to an inland lake. Freshwater Biology **59**:1897-1901.
- Palkovacs, E. P., and D. M. Post. 2008. Eco-evolutionary interactions between predators and prey: can predator-induced changes to prey communities feed back to shape predator foraging traits? Evolutionary Ecology Research **10**:699-720.
- Palkovacs, E. P., and D. M. Post. 2009. Experimental evidence that phenotypic divergence in predators drives community divergence in prey. Ecology **90**:300-305.
- Post, D. M., E. P. Palkovacs, E. G. Schielke, and S. I. Dodson. 2008. Intraspecific variation in a predator affects community structure and cascading trophic interactions. Ecology **89**:2019-2032.
- Schielke, E. G., E. P. Palkovacs, and D. M. Post. 2011. Eco-evolutionary feedbacks drive niche differences in alewives. Biological Theory **6**:211-219.
- Twining, C. W., D. C. West, and D. M. Post. 2013. Historical changes in nutrient inputs from humans and anadromous fish in New England's coastal watersheds. Limnology And Oceanography **58**:1286–1300.
- Walters, A. W., R. T. Barnes, and D. M. Post. 2009. Anadromous alewives (*Alosa pseudoharengus*) contribute marine-derived nutrients to coastal stream food webs. Canadian Journal of Fisheries and Aquatic Sciences **66**:439-448.
- Weis, J. J., and D. M. Post. 2013. Intraspecific variation in a predator drives cascading variation in primary producer community composition. Oikos **122**:1343-1349.
- West, D. C., A. W. Walters, S. Gephard, and D. M. Post. 2010. Nutrient loading by anadromous alewife (Alosa pseudoharengus): contemporary patterns and predictions for restoration efforts. Canadian Journal of Fisheries and Aquatic Sciences **67**:1211-1220.

ORIGINAL ARTICLE





Evaluating the potential for prezygotic isolation and hybridization between landlocked and anadromous alewife (Alosa pseudoharengus) following secondary contact

Katherine A. Littrell¹ | David Ellis² | Stephen R. Gephard² | Andrew D. MacDonald¹ | Eric P. Palkovacs³ | Katherine Scranton⁴ | David M. Post¹

Correspondence

Katherine A. Littrell, Department of Ecology and Evolutionary Biology, Yale University, 165 Prospect Street, New Haven, CT 06511. Email: katherine.littrell@yale.edu

Funding information

Northeast Regional Conservation Needs Grant Program; Yale University Chair's Fund; National Science Foundation Graduate Research Fellowship Program; State Wildlife Grant administered by the Connecticut Department of Energy and Environmental Protection; National Science Foundation, Grant/Award Number: NSF-DEB 1556378 and NSF-DEB 1556848

Abstract

The recent increase in river restoration projects is altering habitat connectivity for many aquatic species, increasing the chance that previously isolated populations will come into secondary contact. Anadromous and landlocked alewife (Alosa pseudoharengus) are currently undergoing secondary contact as a result of a fishway installation at Rogers Lake in Old Lyme, Connecticut. To determine the degree of prezygotic isolation and potential for hybridization between alewife life history forms, we constructed spawning time distributions for two anadromous and three landlocked alewife populations using otolith-derived age estimates. In addition, we analyzed long-term data from anadromous alewife migratory spawning runs to look for trends in arrival date and spawning time. Our results indicated that anadromous alewife spawned earlier and over a shorter duration than landlocked alewife, but 3%-13% of landlocked alewife spawning overlapped with the anadromous alewife spawning period. The degree of spawning time overlap was primarily driven by annual and population-level variation in the timing of spawning by landlocked alewife, whereas the timing and duration of spawning for anadromous alewife were found to be relatively invariant among years in our study system. For alewife and many other anadromous fish species, the increase in fish passage river restoration projects in the coming decades will re-establish habitat connectivity and may bring isolated populations into contact. Hybridization between life history forms may occur when prezygotic isolating mechanisms are minimal, leading to potentially rapid ecological and evolutionary changes in restored habitats.

KEYWORDS

dam removal, fish passage, habitat connectivity, hybridization, prezygotic isolation, secondary contact

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2018 The Authors. Evolutionary Applications published by John Wiley & Sons Ltd

¹Department of Ecology and Evolutionary Biology, Yale University, New Haven, Connecticut

²Fisheries Division, Connecticut Department of Energy and Environmental Protection, Old Lyme, Connecticut

³Long Marine Laboratory, University of California Santa Cruz, Santa Cruz, California

⁴University of California Los Angeles, Los Angeles, California

1 | INTRODUCTION

In the last three centuries, many anadromous fish populations have declined to <1%-10% of their former abundance (Limburg & Waldman, 2009). Dams have historically been a leading contributor to the loss of anadromous fishes worldwide (Hall, Jordaan, & Frisk, 2012; Liermann, Nilsson, Robertson, & Ng, 2012). For anadromous species, dams can block access to high-quality spawning grounds, increase competition for available spawning sites, increase migration time, create lethal water conditions during migration, and decrease offspring survival (Gosset, Rives, & Labonne, 2006; Hall et al., 2012; Harnish, Sharma, McMichael, Langshaw, & Pearsons, 2014; Locke, Hanson, Klassen, Richardson, & Aubé, 2003; Zhou, Zhao, Song, Bi, & Zhang, 2014). The multiple negative impacts of dams, combined with other anthropogenic stressors, have contributed to the local extirpation of many anadromous fish species (Beamish & Northcote, 1989; Gao, Lin, Li, Duan, & Liu, 2016; Rolls, Stewart-Koster, Ellison, Faggotter, & Roberts, 2014). To reverse these impacts, restoration projects are removing dams or installing fishways to restore access to spawning sites. Over 1,300 dam removals and hundreds of fish passage projects have been completed in the United States within the last several decades (Lenhart, 2003; Rivers, 2017; U.S. Fish & Wildlife Service, 2012). Fishway installations and dam removals are completed with the intent of restoring anadromous spawning migrations, but there is little consensus on what constitutes successful restoration and there can be unforeseen impacts on population recovery and local ecosystems (Babbitt, 2002; Bernhardt et al., 2007; Grant, 2001; Hart et al., 2002).

One consequence of restoring habitat connectivity is the increased probability that historically separated populations will come into contact with each other. Such a scenario represents an example of secondary contact, when two evolutionary diverged lineages come into contact after a period of geographic isolation (Hutchings & Myers, 1985; Jones, Brown, Pemberton, & Braithwaite, 2006; Tulp et al., 2013). The ecological and evolutionary processes that occur at the very onset of secondary contact can be complex and determine whether populations undergo speciation, fuse into a single population, or create a hybrid zone (Barton & Hewitt, 1989; Coyne, 1992; Hewitt, 1988). Habitat restoration efforts may provide opportunities to study these processes in natural populations. As restoration efforts seek to reconnect fragmented ecosystems (Baguette, Blanchet, Legrand, Stevens, & Turlure, 2013; Beninde, Veith, & Hochkirch, 2015; Resasco, Bruna, Haddad, Banks-Leite, & Margules, 2017), instances of secondary contact are likely to become more common. Studying secondary contact as it occurs can provide valuable insight into the biological mechanisms that generate and maintain biodiversity in a wide range of taxa and inform conservation efforts that strive to restore ecosystem connectivity.

Anadromous fishes are an excellent study system for questions pertaining to secondary contact, as their diverse life history forms can become genetically isolated via natural or anthropogenic mechanisms. For example, dam construction can isolate a portion of an anadromous fish population in freshwater, leading to the evolution of an alternate life history that completes its entire lifecycle in freshwater

(Berg. 1985: Palkovacs, Dion. Post. & Caccone, 2008: Pearse et al., 2009). The evolution of freshwater resident life history forms is found in numerous taxonomic families, including Salmonidae, Osmeridae, Clupeidae, Gasterosteidae, Petromyzontidae, and Plecoglossidae (McDowall, 1988). Examples of adaptations documented in freshwater resident fish populations include smaller size at maturity, smaller foraging morphology (Jones, Palkovacs, & Post, 2013; Karve, von Hippel, & Bell, 2008; Palkovacs & Post, 2009), alternate reproductive strategies (Campbell, 1977; Closs, Hicks, & Jellyman, 2013; Gulseth & Nilssen, 2001), and a decreased tolerance to salinity (Velotta, McCormick, O'Neill, & Schultz, 2014; Velotta, McCormick, & Schultz, 2015). Taken together, this suite of life history adaptations creates the potential for considerable ecological and evolutionary differentiation between freshwater resident and ancestral anadromous forms. In cases where there are significant morphological, ecological, or genetic differences between populations, secondary contact could result in competition and competitive exclusion (Perry, Feder, Dwyer, & Lodge, 2001), coexistence through niche partitioning and character displacement (Aguilee, de Becdelievre, Lambert, & Claessen, 2011; Levine & HilleRisLambers, 2009; Mayfield & Levine, 2010), speciation via reinforcement (Hasselman et al., 2014), or hybridization (Barton & Hewitt, 1985, 1989; Hewitt, 1988). Secondary contact that results from restoring spatial connectivity could have large impacts on anadromous fish populations, but the outcome depends on the potential for gene flow between anadromous and freshwater resident populations. Gene flow between populations is possible if prezygotic isolating mechanisms such as divergent mating behavior, spatial isolation, temporal isolation, and gamete incompatibility are minimal (Coyne & Orr, 2004).

Species in the genus Alosa (shad and river herring) have been identified as the second most prevalent genus in threatened freshwater ecoregions, with 14 of 25 alosine species located in heavily obstructed river systems (Bernhardt et al., 2005; Vörösmarty et al., 2010). In North America, alewife (Alosa pseudoharengus) is the target of conservation management plans along the Atlantic coast using fishway installations to restore migratory pathways to historical spawning grounds (Hasselman & Limburg, 2012; Lake, Ravana, & Saunders, 2012). Many of the systems targeted for restoration contain independently evolving populations of landlocked alewife, the freshwater resident form of anadromous alewife, in lakes and reservoirs above the dams (Palkovacs et al., 2008). In lakes in southern Connecticut, landlocked alewife were isolated from anadromous populations 300-500 years ago, likely as a result of colonial dam construction (Palkovacs et al., 2008; Twining & Post, 2013). Landlocked alewife populations have rapidly evolved ecological and evolutionary differences from anadromous alewife during this relatively short period of reproductive isolation (Jones et al., 2013; Palkovacs, Mandeville, & Post, 2014; Palkovacs & Post, 2009; Post, Palkovacs, Schielke, & Dodson, 2008; Schielke, Palkovacs, & Post, 2011).

Here, we ask how variation in spawning time might influence the potential for gene flow between landlocked and anadromous alewife populations. Alewife can hybridize with its sister species, blueback herring (*Alosa aestivalis*) (Hasselman et al., 2014). This suggests a lack of postzygotic isolating mechanisms and a high probability of

hybridization between life history forms if there is spawning time overlap. Alewife is a broadcast spawner and does not exhibit complex behaviors during mating that may impede hybridization. In general, anadromous alewife has been reported to spawn earlier (April–June) (Kissil, 1974) than landlocked alewife (May–August) (Nigro & Ney, 1982). Therefore, divergence in spawning time may create prezygotic isolation, which could be an important limiting factor for hybridization. We used otolith-based age estimates and migration data to quantify the timing of reproduction and explore sources of variation in spawning period that may contribute to or inhibit hybridization between alewife life history forms.

2 | MATERIALS AND METHODS

2.1 | Data collection

2.1.1 | Migration time

We acquired adult anadromous alewife migration data from three fishways in Connecticut to examine historical trends in anadromous alewife spawning behavior. The Connecticut Department of Energy and Environmental Protection (CT DEEP) collected the data across multiple years. The fishways were located in the Branford Supply Ponds (2006-2016), Mill Brook leading into Rogers Lake (2002-2016), and Bride Brook leading into Bride Lake (2003-2016). Daily fish counts of adult anadromous alewife passing through the fishway into the lake and water temperature were collected by CT DEEP 5-7 days a week and recorded until spent adult fish began returning to sea, at which point the fish counters were removed. Due to the removal of the fish counter and termination of fish counts with outmigration of adults, we excluded the upper and lower 5% of the run, constraining the data to the middle 90% of the run (based on the total number of fish passing through the fishway). We acquired daily ocean surface temperature data (New London, CT) from NOAA's

National Centers for Environmental Information water temperature database (NOAA Tides and Currents, 2018).

2.1.2 | Spawning time

We estimated alewife spawning time in five lakes in southern Connecticut. All of the systems sampled are within 25 km of the coast (see Palkovacs et al. (2008) for a map of the study sites). Bride Lake (41.3276°N, 72.2378°W) and Dodge Pond (41.3275°N, 72.1986°W) are spawning grounds for anadromous alewife and support young-of-the-year (YOY) alewife for the period of time from hatching in the spring to marine migration in the fall. Pattagansett (41.3728°N, 72.2312°W) and Quonnipaug (41.3889°N, 72.6986°W) Lakes are populated by landlocked alewife, with access from the ocean blocked by dams. Rogers Lake (41.3637°N, 72.3000°W) is populated by landlocked alewife, but a recent fishway installation in 2014 and anadromous alewife stocking program reintroduced anadromous alewife into Rogers Lake. Thus, Rogers Lake is the site of ongoing secondary contact between alewife life history forms. A future secondary contact event is likely to occur in Pattagansett Lake, which is under consideration for a fishway installation within the coming decade. Pattagansett Lake and Dodge Pond are within the Pattagansett River watershed and anadromous alewives genetically similar to those in Dodge Pond may eventually populate Pattagansett Lake.

Fish were captured from all five lakes between July 27 and August 21 (2013, 2014, 2015) using a small research purse seine (4.87 m deep $\times 35.36$ m long, mesh size 1/16 inches) designed to encircle 100 m². We collected 50–150 fish in a single night from the pelagic and littoral zones in six separate seine sets performed in different locations across the lake (Table 1). All fish were immediately euthanized using MS-222 and stored at -20° C until processing. To quantify the potential for prezygotic isolation between anadromous and landlocked alewife, we used otolith-derived age estimates to

TARIF 1	Alewife campling	information and	age estimation	licing a	length-age regression

Lake	Year	Sampling date	Fish total length (mm)	Fish aged by otoliths	Fish aged by regression	Length-age regression equation	R^2
Bride	2014	5 Aug	33-68	104	0	y = 47.81 + 0.579x	0.28
Bride	2015	4 Aug	35-66	100	0	y = 55.28 + 0.349x	0.14
Dodge	2014	4 Aug	31-55	100	0	y = 41.89 + 0.834x	0.26
Dodge	2015	3 Aug	30-54	99	0	y = 45.30 + 0.731x	0.25
Pattagansett	2014	7 Aug	25-85	79	9	y = 2.69 + 0.904x	0.89
Pattagansett	2015	6 Aug	33-82	102	13	y = -4.20 + 1.067x	0.87
Quonnipaug	2014	21 Aug	18-91	100	65	y = 11.19 + 0.712x	0.89
Quonnipaug	2015	7 Aug	16-85	100	15	y = 5.52 + 0.845x	0.93
Rogers	2013	13 Aug	24-63	49	71	y = 12.40 + 0.882x	0.62
Rogers	2014 ^a	6 Aug	11-75	21	51	y = 12.40 + 0.882x	0.62
Rogers	2015	27 July, 5 Aug	20-79	99	286	y = 5.13 + 0.894x	0.82

^aThere were too few otoliths from the Rogers 2014 sampling season to accurately estimate age. We used the length-age regression from 2013.

develop spawning time distributions for all five lakes (two anadromous, three landlocked populations). For Rogers Lake, no adult anadromous alewife passed through the fishway into the lake in 2014. In the spring of 2015, Rogers Lake was stocked with 130 adult anadromous alewife from Mill Brook. Based on this very small number of adult anadromous alewife from Mill Brook, we believe it is unlikely that we captured YOY anadromous alewife in 2015. Therefore, we treated Rogers Lake in 2014 and 2015 as purely landlocked populations. To supplement the anadromous alewife population in Rogers Lake, adult anadromous alewife from Bride Lake have been stocked into Rogers Lake each year starting in 2016.

We estimated alewife hatch date by counting otolith daily growth increments. Both sagittal otoliths were removed and mounted on a glass microscope slide with a heat malleable plastic resin. Only 20 otoliths were available from Rogers Lake in 2014. We used the length-age regression from the Rogers Lake 2013 population to estimate age in 50 additional Rogers Lake 2014 fish. We polished opaque otoliths using 1,000 and 2,000 grit automotive sanding paper and fine polished the surface using a $0.05 \, \mu m$ slurry of alumina powder. Increments were counted a total of five times by the same observer under 20x to 40x magnification (Leica DM LS2). We excluded samples that were inconsistent across counts, typically with differences >5-7 days between individual counts. Approximately 100 random fish per lake, including the largest and smallest individuals, were counted to capture the full range of the spawning period. In some years, <100 fish were available due to low capture rates in the field and damage to the otoliths during extraction, particularly in fish <20 mm. For landlocked fish, there was a strong relationship between length and age (Table 1). We used the age-length regression equation for each lake and year to estimate the age of additional fish and replace fish with damaged otoliths. Only 20 otoliths were available from Rogers Lake in 2014. We used the length-age regression from the Rogers Lake 2013 population to estimate age in 50 additional Roger Lake 2014 fish. Anadromous fish did not exhibit a strong relationship between length and age; therefore, we did not use length-age regressions to estimate hatch date in additional fish.

Spawning date was calculated by combining data on otolith-derived hatch dates with temperature-derived estimates of growth rate during development within the egg. We used temperature profiles from biweekly water sampling at the deepest part of each lake to interpolate daily water temperatures throughout the spawning season. The first 3 m of the epilimnion was averaged to approximate the temperature in benthic habitat where alewife spawn. Our formula for calculating development time in the egg from lake temperatures, $T = 114.05e^{-0.048F}$ $(R^2 = 0.94)$, was derived by digitization of the data presented in figure 3 of Edsall (1970), where T is days to hatching and F is temperate in degrees Fahrenheit. The egg development time was added to our otolithderived hatch times to determine days since spawning. The number of days since spawning was subtracted from the date of capture to calculate a spawning date for each fish. All fish were collected under CT DEEP Scientific Collector's Permits SC-11016 and SC-14023 and handled in compliance with Yale's Institutional Animal Care and Use Committee protocols 10734 (2012) and 10734 (2015).

2.2 | Statistical analysis

2.2.1 | Migration data

As the anadromous alewife spawning migration (movement upriver) occurs in intermittent pulses over the spring, we isolated several metrics as indicators of the overall timing of migration: the day on which 10%, 50%, or 90% of the population had migrated and the day of peak (maximum) migration. We used a linear regression with Gaussian distributed errors (Bates, Mächler, Bolker, & Walker, 2015; R Core Team, 2017) to estimate the relationship between the timing of migration and ocean water temperature, while accounting for any trend across years and individual population effects. We treated all factors as fixed effects, including the differences between populations because of the low replication of rivers (n = 3). We also included a second-order ocean temperature term that created a quadratic relationship with a single peak for migration. This term allowed for the possibility of a thermal optimum for migration, where migration would decrease at warmer or colder temperatures. We performed model selection using AIC to determine the order of the relationship of migration with ocean temperature and whether any covariates explained the variation in the relationship between migration and ocean temperature. We also used a simple linear regression (Bates et al., 2015; R Core Team, 2017) to determine whether the size of the migratory run (the total number of fish observed) predicted the overall duration of the migration period.

2.2.2 | Spawning data

Anadromous and landlocked alewife spawning occurs from April to mid-July, creating a set of unimodal distributions of spawning dates that differ by alewife life history form, lake, and year. We modeled the distribution of spawning dates using time-to-event analysis, commonly used in survival analysis. Time-to-event models assume some inherent variability in the timing of the event; individuals experience the event at times distributed according to a probability distribution (Hougaard, 2000). The instantaneous probability of the event (the hazard function) can be affected by covariates making individuals with certain traits or in certain groups more or less likely to experience the event earlier or later.

We generated event (spawning) times for each individual, defined as the time from January 1 to the estimated spawning date. As the resolution of our estimates of spawning time was on the order of a day, we used interval censoring to account for the fact that spawning may have occurred at any time during the day (but taking the exact event time as the midpoint of the day did not affect our results). We assumed spawning times followed a Weibull distribution with shape and scale parameters that allow flexibility in the shape of the underlying instantaneous probability of spawning (hazard function). We hypothesized that spawning may be affected by alewife life history form, while accounting for differences between years and between lakes.

We considered lake temperature as a factor, but water temperature in our five lakes increased linearly over the summer, making it impossible to separate the effects of time from the effects of water temperature (Figure A1 in Appendix S1). Some alewife populations spawned earlier in cooler water, and some spawned later in warmer water. Water temperatures in the five lakes were very similar, and landlocked lakes were not more similar to each other than they were to anadromous lakes. Thus, we excluded lake temperature from our analysis, supported by the fact that lakes varied only very slightly, and not systematically, in temperature (see Appendix S1 for more detail).

We approximated and maximized the likelihoods in R with original code (R Core Team, 2017) and performed model selection with AIC (Appendix S1). This let us estimate the parameters of the distribution of spawning times (the shape and rate parameters) as well as the coefficients of the covariates: alewife life history form, lake, and year. From these estimates, we calculated the mean and variance of the estimated spawning time distributions, as well as the probability of both life history forms spawning at the same time (Appendix S1). We estimated the potential for hybridization using two metrics: the percent of the population expected to experience an "interbreeding event" and the "spawning overlap." The percent of the population expected to experience an interbreeding event was derived from the cumulative probability that one anadromous and one landlocked alewife spawn on the same day throughout the entire spawning season. Spawning overlap was defined as the percentage of the landlocked population that spawns at any point in time within the anadromous spawning distribution, but not necessarily on the same day. Neither

measure of hybridization potential makes any assumptions about sex ratios or spawning behavior. Anadromous populations were treated as a single "anadromous" spawning time distribution due to their nearly identical distributions and for comparisons with landlocked populations. The Rogers Lake and Quonnipaug Lake landlocked populations were also treated as one spawning time distribution because they did not differ statistically from one another. We compared the Pattagansett Lake landlocked spawning time distributions to the anadromous distribution independently because they were statistically different from the other two landlocked alewife populations in Rogers Lake and Quonnipaug Lake.

3 | RESULTS

3.1 | Migration

Adult anadromous alewife generally migrated from mid-March to mid-May, with anadromous alewife starting to arrive on the spawning grounds in small numbers as early as late February (Figure 1). The majority of the anadromous alewife migration occurred within a 20- to 30-day period beginning the second week of April. Across the four migratory timing metrics, the models that were highly supported included ocean surface temperature and migratory river, indicating that there was a significant relationship of migration to ocean temperature, but that it varied across populations. Models

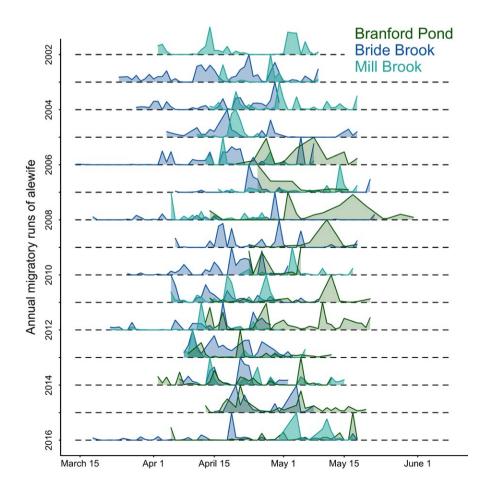


FIGURE 1 Probability distributions of adult anadromous alewife migration by arrival date from 2002 to 2016 in Bride Brook, Mill Brook, and Branford Supply Ponds. The probability distributions are based on CT DEEP counts at each location using electronic fish counters. The annual number of anadromous alewife passing through each fishway is provided in Table B1 in Appendix S2

with a second-order term for ocean temperature were equally as likely as those with a linear relationship, indicating no support for a thermal optimum for migration (Appendix S3). We found no evidence for a secular change in the timing of migration; year was not a factor in the most highly supported models. Although not significant, the anadromous alewife run at Bride Brook tended to arrive 15–20 days earlier than anadromous alewife in Mill Brook and Branford Supply Ponds. Anadromous alewife migrated when ocean surface temperatures were between 4.9 and 16.0°C, with average migration temperatures between 7.0 and 12.0°C.

Run size (number of spawning adults) at Bride Brook over the last decade ranged from 68,731 to 354,862 fish, with a mean of 162,075 \pm 94,588 fish. The Mill Brook had runs ranging from 99 to 15,362 fish (mean = 5,620 \pm 5,333 fish), and the run at the Branford Supply Ponds ranged from 563 to 50,668 fish (mean = 8,297 \pm 17,179 fish). Run size negatively correlated with run duration at Bride Brook ($F_{1,11}$ = 9.518, p = 0.01, R^2 = 0.4151), but there was no relationship between run size and run duration at Mill Brook or the Branford Supply Ponds, likely due to smaller run sizes.

3.2 | Spawning

Landlocked alewife spawned later and over a longer duration than anadromous alewife. Landlocked alewife also had greater variation in spawning date than anadromous alewife (Figure 2). Life history form was the most important explanatory variable for spawning time in alewife, but spawning time did differ by year and population

(Table 2). The Rogers Lake landlocked population spawned much earlier in 2013 than in 2014 and 2015. The Pattagansett Lake population spawned earlier than all other landlocked populations in both 2014 and 2015.

Anadromous alewife spawned over a period of 35-40 days, beginning as early as April 26 and continuing until the first week of June. The water temperature during this time period ranged from 12.5 to 21.2°C. Mean spawning date for anadromous alewife was May 19 ± 9.3 days (Table 3). Peak spawning occurred in mid-May when water temperatures reached 17.0-20.0°C. There were subtle differences in the spawning periods of anadromous populations. with Dodge Pond alewife initiating and reaching peak spawning up to a week earlier than alewife in Bride Lake. Although spawning duration was nearly identical in both populations, in Dodge Pond 2014, we detected one fish that hatched in late June. This individual extended the estimated spawning period from 38 to 59 days; this was the only fish in the sample that hatched after 2nd June. We detected a 2-week delay between the end of migration in Bride Lake and peak spawning activity, which was similar to other anadromous alewife populations (Rosset et al., 2017).

Landlocked alewife populations started spawning in early to late May and continued spawning for approximately 60 days, when water temperatures were between 13.5 and 27.5°C. The Pattagansett Lake and Quonnipaug Lake populations commenced spawning between May 2 and May 15, while the Rogers Lake population started spawning slightly later from May 11 to May 20 depending on the year. The Pattagansett Lake population reached peak spawning in mid-June

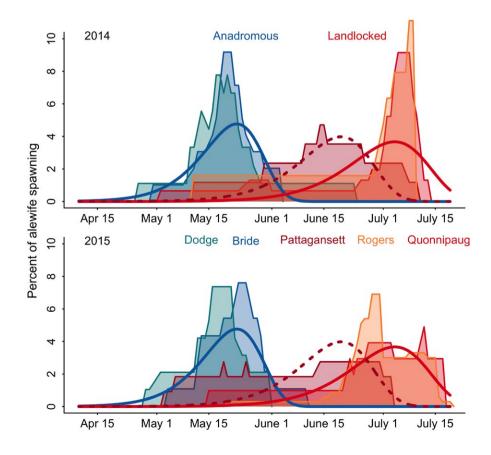


FIGURE 2 Probability distributions of spawning time for Bride/Dodge anadromous (blue solid line), Rogers/Quonnipaug landlocked (red solid line), and Pattagansett landlocked (red dotted line) alewife from 2014 to 2015. The colored probability distributions represent otolith-derived spawning dates for each population

TABLE 2 Models and AIC selection scores for alewife spawning time

Model	Delta AIC
y ~ form ^a + 2013 + all lakes	0
y ~ form + all years + Pattagansett Lake	0.466
y ~ form + 2013 + Pattagansett Lake	2.24
y ~ form + all years + all lakes	20.5
y ~ form + Pattagansett Lake	173
y ~ form + 2013	261
y ~ form + all years	263
y ~ form	391
y ~ form + all lakes	749
y ~ all years + all lakes	765
y ~ 1	2,057

^aForm = alewife life history form.

TABLE 3 Mean spawning date for anadromous (A) and landlocked (L) alewife populations

Population	Mean spawning date	SD (days)
2014/2015 Bride and Dodge (A)	May 19	9.3
2014/2015 Rogers and Quonnipaug (L)	June 29	12.1
2014/2015 Pattagansett (L)	June 15	11.1
2013 Rogers (L)	June 15	11.1

(22.0–25.0°C), 2–3 weeks earlier than all other landlocked lakes. Peak spawning in Quonnipaug Lake occurred in early to mid-July (25.0–27.0°C). In the Rogers Lake 2014 and 2015 populations, peak spawning also occurred from early to mid-July (25.0–27.0°C), but peak spawning occurred in mid-June during the 2013 spawning season (22.0–24.0°C).

We detected low, but variable levels of spawning time overlap and interbreeding events between anadromous and landlocked alewife. Our model-derived spawning overlap estimate, considering all populations and years, was <15%. The degree of overlap varied by year and population, with some landlocked populations in certain years overlapping more with anadromous alewife than others. Approximately 3% of landlocked alewives in Rogers Lake and Quonnipaug Lake were predicted to have spawned within the anadromous alewife spawning period. Spawning overlap was higher in Pattagansett Lake at 13%. Our data typically followed the model with the exception of the Pattagansett Lake 2015 population. In 2015, the data for Pattagansett Lake deviated from our spawning model and indicated that spawning overlap with anadromous alewife was as high as 30%. The spawning overlap estimates were relatively liberal, as they did not consider what day both alewife life history forms spawned. A more conservative estimate of the hybridization potential was the percentage of the landlocked population that experienced interbreeding events, or the proportion of the population that spawned on the same day as anadromous alewife in a single spawning season. Our model indicated that 0.1% of the Rogers/Quonnipaug Lake landlocked population and 0.4% of the Pattagansett Lake landlocked population spawned on the same day as anadromous alewife. In both landlocked populations, interbreeding events with anadromous alewife were most likely to occur during the last week of May.

4 | DISCUSSION

Re-establishing ecosystem connectivity, which may bring historically isolated populations into secondary contact, has become a common objective in conservation and restoration planning. For anadromous fish, increasing habitat connectivity as a result of dam removals and fishway installations will bring anadromous populations into secondary contact with previously isolated and independently evolving landlocked populations. Variation in reproductive timing between anadromous and landlocked populations will influence the potential for hybridization and level of introgression between life history forms during contact. For alewife in our study lakes, the probability of concurrent spawning between life history forms was low, but variable, due to strong temporal differences in spawning behavior. Landlocked alewife reached peak spawning 1.5 months later than anadromous alewife, but the landlocked alewife spawning distributions did overlap with the anadromous alewife spawning distribution, with 3%-13% of landlocked alewife spawning during the anadromous alewife spawning period. For the Rogers Lake landlocked alewife population, which is experiencing secondary contact with anadromous alewife from Bride Lake, 0.1% of landlocked alewife spawned on the same day as anadromous alewife. Pattagansett Lake landlocked alewife will experience secondary contact with anadromous alewife genetically similar to alewife in Dodge Pond after a fishway installation in the next decade. Due to earlier spawning, 0.4% of Pattagansett Lake landlocked alewife spawned on the same day as anadromous alewife from Dodge Pond. Our model indicated that 13% of the Pattagansett Lake landlocked alewife population spawned within the anadromous alewife spawning period, but the data suggested that this estimate may be as high as 30%. Spawning time overlap and the potential for hybridization between life history forms are largely dependent on the spawning behavior of the particular landlocked populations under consideration. We anticipate that anadromous populations may be more similar in their spawning behavior due to a shared marine environment and similar migratory constraints, while landlocked spawning patterns are subject to local selection pressures and free to diverge from one another across lakes. Our results support this idea.

Our anadromous alewife spawning dates and temperatures are comparable to that of other anadromous alewife populations along the Atlantic coast (Rosset et al., 2017). The anadromous alewife population at Bride Lake has been studied repeatedly, with a focus on the timing of adult anadromous alewife migration. Kissil (1974) observed anadromous alewife migrating from March to June when stream temperatures reached 4.0–5.0°C and adults remained in the

lake to spawn anywhere from 3 days to 3 months after arrival. Ellis and Vokoun (2009) identified a stream temperature of 9.0°C as indicative of the start of migration and 13.0°C as the strongest predictor of average run time in three Connecticut anadromous alewife populations, including Bride Lake. Our data indicated that anadromous alewife migrated from March to June when stream temperatures were between 6.0 and 22.0°C. Peak migration occurred from mid-April to mid-May when stream temperatures averaged between 11.0 and 14.0°C. Anadromous alewife spawned in Bride Lake and Dodge Pond in the last week of April and peak spawning occurred in mid-May when lake temperatures reached 17.0-20.0°C. We did not detect the effect of climate change on advancing the arrival date of adult anadromous alewife in Connecticut predicted by Ellis and Vokoun (2009), nor did we identify the Bride Brook population as arriving significantly earlier than other anadromous alewife populations. We note that our results differ because we restricted our dataset to the last 15 years and did not use the weighted-mean migration temperature in our analysis. We also used the first 5% of the cumulative run total to define run initiation.

Landlocked alewife populations along the Atlantic coast and Laurentian Great Lakes region spawn primarily in late May through July (Gross, 1959; Lackey, 1970; Odell, 1934; Rothschild, 1966), but the spawning season is variable and can extend into August (Pritchard, 1929). Landlocked alewife in Claytor Lake, Virginia, spawned from early May once water temperatures reached 16.0-18.0°C, until early August when temperatures reached 24.0-27.0°C. Peak spawning occurred in the third week of May when temperatures were between 21.0 and 23.0°C (Nigro & Ney, 1982). Nigro and Ney (1982) noted that during this 1974 spawning season, the Claytor Lake landlocked alewife population spawned a month earlier and 4-9 weeks longer than northern landlocked alewife populations. We observed similar spawning patterns to other northern lakes in Rogers, Quonnipaug, and Pattagansett Lakes. For most lakes and years, landlocked alewife spawning started in mid-May and continued through July at temperatures between 13.5 and 27.5°C. Peak landlocked alewife spawning occurred when temperatures reached 22.0-27.0°C.

The difference in spawning time between anadromous and landlocked alewife may be driven by selective constraints related to migration. Anadromous alewives mature along the Atlantic coast several hundred kilometers or more from their spawning grounds (Neves, 1981). Alewife, as with other anadromous species, relies on internal and external cues at sea to correctly time migration to maximize its survival en route to spawning sites (Berdahl, Westley, Levin, Couzin, & Quinn, 2016; Hansen, Jonsson, & Jonsson, 1993; Quinn & Adams, 1996). Anadromous fishes face the added challenge of timing migration to align with peak conditions for offspring survival. Ideal conditions for migration and spawning may not coincide; a trade-off may exist between ideal timing of migration and spawning that determines the actual timing of reproduction (Quinn, McGinnity, & Reed, 2016). Environmental conditions such as water temperature and stream discharge have been correlated with migratory timing in anadromous species, particularly salmonids (Hodgson

& Quinn, 2002; Quinn & Adams, 1996; Quinn, Hodgson, & Peven, 1997). Spawning sites can become inaccessible or travel conditions lethal during periods of low stream discharge and high temperatures (Goniea et al., 2006; Rand et al., 2006). Iteroparous anadromous species suffer the added constraint of successfully migrating back to sea after spawning. In contrast, landlocked fish are present in the spawning habitat year round, releasing them from many of the selective constraints associated with migration. Landlocked alewife may closely track environmental cues and time spawning to coincide with peak conditions for offspring survival (Lyons et al., 2015). This time period may be much later, and at warmer temperatures, than the period safe for the migration of anadromous alewife.

A hypothesis for the observed differences in spawning time variability between anadromous and landlocked alewife is there are differences in temperature variability between marine and freshwater ecosystems. Oceans exhibit less temperature variability through time than small bodies of freshwater on continents (Cyr & Cyr, 2003). Landlocked alewives have to be adaptable to rapidly fluctuating environmental conditions in comparison with anadromous alewives in a more constant marine growth environment, which ultimately influences sexual maturation, migration, and spawning (Friedland, 1998; Gardner, 1976).

Our data indicated that changes in spawning time overlap between alewife life history forms were caused by yearly and population-level variation in landlocked spawning behavior. The sources of this variation are unknown, as we did not detect any significant environmental differences, most notably lake temperature, between our three landlocked lakes. Instead, genetic differences between populations, resulting from natural selection or genetic drift, could drive spawning time variability. All three of our landlocked lakes are genetically isolated and have evolved independently from each other (Palkovacs et al., 2008). Our landlocked alewife populations are as genetically divergent from each other as they are from their ancestral anadromous alewife populations (Palkovacs et al., 2008). Unlike anadromous alewife populations, which exchange genes over a broad marine region (Palkovacs et al., 2014) and are only subject to selection in freshwater lakes for a few months a year, landlocked alewife populations are genetically isolated in freshwater lakes year round and free to adapt to local conditions (Jones et al., 2013; Palkovacs et al., 2014; Palkovacs & Post, 2009; Post et al., 2008; Schielke et al., 2011).

Our data on spawning date may be affected by in-lake YOY mortality and the outmigration of anadromous alewife. Although we included both the largest (oldest) and smallest (youngest) alewife in each sample, we may have missed a portion of the anadromous YOY cohort that migrated from the lake starting in early June; therefore, it is possible that the anadromous alewife spawning period starts earlier than indicated. We also did not correct for in-lake mortality in either population and limited our sampling to a single date for each lake per year. As a result, the peak spawning time for both life history forms may be earlier than estimated. We believe that the region of spawning time overlap between life history forms, comprised of the late portion of the anadromous

alewife spawning distribution and early portion of the landlocked alewife spawning distribution, is unlikely to be strongly affected by any sampling bias. Our estimated probabilities of spawning overlap may be underestimates, but unlikely to be overestimates, of true spawning potential; what we provide is a conservative estimate of hybridization potential.

The degree of prezygotic isolation between landlocked and anadromous alewife will influence the genetic and ecological outcomes of secondary contact after fish passage river restoration projects. We detected low levels of spawning time overlap between anadromous alewife and two of our landlocked alewife populations (Rogers Lake and Quonnipaug Lake). Less than 3% of landlocked fish in these two lakes spawned at the same time as anadromous alewife. However, among-lake and among-year variation in landlocked alewife spawning patterns can increase spawning time overlap to 13%. Our data from Pattagansett Lake in 2015, which deviated from our model, suggested that this upper estimate may be as high as 30%. A spawning time overlap of 3%-13% provides some potential for hybridization and introgression between life history forms. Our spawning time models do not include any behavioral changes that may increase the hybridization potential between anadromous and landlocked alewife populations. It is possible that the presence of large fertile anadromous females and anadromous spawning activity may trigger landlocked alewife to reproduce earlier than expected and increase spawning time overlap higher than predicted in this analysis (Hobbs, Munday, & Jones, 2004).

We cannot predict how hybridization will impact alewife populations long term. Possible outcomes of hybridization include the introduction of maladaptive genes (Glover et al., 2017), production of competitively superior hybrids (Perry et al., 2001), formation of polymorphic populations (Riva-Rossi, Pascual, Babaluk, García-Asorey, & Halden, 2007), and speciation via character displacement (Hasselman et al., 2014). It is possible that the introduction of maladaptive traits from landlocked populations (Velotta, McCormick, Jones, & Schultz, 2018; Velotta et al., 2014, 2015) into anadromous populations could reduce anadromous alewife migration success and fitness in marine ecosystems. Changes in alewife morphology and behavior, however, can have immediate effects on freshwater ecosystems. Both alewife life history forms are known for their strong and divergent roles in shaping zooplankton community structure in freshwater lakes. The divergent zooplankton communities are the result of differences in alewife life history (specifically freshwater residency time of YOY), foraging behavior, and morphology (Jones et al., 2013; Palkovacs & Post, 2009; Post et al., 2008). Hybridization between anadromous and landlocked alewife may result in offspring with intermediate size, foraging morphology, and mixed migratory strategies. Due to the strong ecological effects of alewife on freshwater zooplankton community structure, differences in traits among hybrid alewife have the potential to cause direct and rapid changes in the ecology of freshwater lakes.

There are many anadromous fish species facing changes in habitat connectivity as fish passage projects accelerate in the coming decades (Pohl, 2002). Many of these species also have isolated

landlocked populations behind migratory barriers (Apgar, Pearse, & Palkovacs, 2017; Berg, 1985; Palkovacs et al., 2008; Pearse et al., 2009), and it is unknown how secondary contact between life history forms will affect populations of conservation concern. When prezygotic isolation prevents gene flow, it is possible for speciation to occur. In contrast, gene flow may result in complex outcomes ranging from the fusion of populations to the formation of a stable hybrid zone (Barton & Hewitt, 1989; Coyne, 1992; Hewitt, 1988). Ecological interactions, such as competition, combined with introgression between life history forms, may have significant implications for the evolution, ecology, and management of anadromous fish populations during secondary contact with freshwater resident populations.

Little is known about the success of all types of river restoration projects or how they affect ecosystems due to a lack of long-term monitoring (Babbitt, 2002; Bernhardt et al., 2007; Grant, 2001; Hart et al., 2002). Pressing questions include identifying the historical impacts of dams on ecosystems and predicting the impact of dam removals or fish passage projects on future ecosystem function. Part of the ecological response will involve evolutionary processes, including secondary contact. In this manner, fish passage river restoration projects are an ecosystem-level experiment to test how changing habitat connectivity impacts ecological and evolutionary dynamics.

Restoration projects that aim to re-establish landscape connectivity may commonly drive secondary contact between historically isolated populations in a variety of taxa. Similar to fishway installations and dam removals in aquatic ecosystems, the construction of wildlife corridors to connect isolated populations in terrestrial ecosystems also alters habitat connectivity (Beier & Noss, 1998). Adaptation to different ecological conditions in isolated habitats is likely to result in the evolution of divergent traits (Bicudo, Anciães, Benchimol, Peres, & Simões, 2016; Fraser, Debes, Bernatchez, & Hutchings, 2014; Santos & Araújo, 2015; Zastavniouk, Weir, & Fraser, 2017). Trait differences between populations can lead to ecological and evolutionary interactions after secondary contact that have the potential to influence the outcome of restoration and conservation projects. Changes in habitat connectivity are not limited to restoration projects. Anthropogenic changes to species ranges (Potts et al., 2014), the spread of invasive species (Kovach et al., 2015; Perry et al., 2001; Zaccara, Antognazza, Buonerba, Britton, & Crosa, 2014), and pollution (Seehausen, van Alphen, & Witte, 1997) are breaking down reproductive barriers and bringing previously isolated populations or closely related species into secondary contact. Using restoration projects as a model to study the ecological and evolutionary dynamics of secondary contact will help inform future conservation and restoration efforts as anthropogenic changes to habitat connectivity accelerate in the coming decades.

ACKNOWLEDGEMENTS

We would like to thank Dr. Karin Limburg for her time demonstrating alewife otolith extraction and aging. Funding was provided by the

National Science Foundation (NSF) Graduate Research Fellowship Program; Yale University's Chair's Fund; NSF (NSF-DEB #1556378, NSF-DEB # 1556848); the Northeast Regional Conservation Needs Grants Program; and the State Wildlife Grant administered by the Connecticut Department of Energy and Environmental Protection. This publication does not express the views of the Connecticut Department of Energy and Environmental Protection or the State of Connecticut. The views and opinions expressed are those of the authors.

CONFLICT OF INTEREST

None declared.

DATA ACCESSIBILITY

Data for this study are available from the Dryad Digital Repository: https://doi.org/10.5061/dryad.7qt4220.

ORCID

Katherine A. Littrell http://orcid.org/0000-0001-8113-2408

Eric P. Palkovacs http://orcid.org/0000-0002-5496-7263

Katherine Scranton http://orcid.org/0000-0002-7656-676X

David M. Post http://orcid.org/0000-0003-1434-7729

REFERENCES

- Aguilee, R., de Becdelievre, B., Lambert, A., & Claessen, D. (2011). Under which conditions is character displacement a likely outcome of secondary contact? *Journal of Biological Dynamics*, 5, 135–146. https://doi.org/10.1080/17513758.2010.491559
- Apgar, T. M., Pearse, D. E., & Palkovacs, E. P. (2017). Evolutionary restoration potential evaluated through the use of a trait-linked genetic marker. *Evolutionary Applications*, 10, 485–497. https://doi.org/10.1111/eva.12471
- Babbitt, B. (2002). What goes up, may come down. BioScience, 52,656-658. https://doi.org/10.1641/0006-3568(2002)052[0656:WGUMCD] 2.0.CO;2
- Baguette, M., Blanchet, S., Legrand, D., Stevens, V. M., & Turlure, C. (2013). Individual dispersal, landscape connectivity and ecological networks. *Biological Reviews*, 88, 310–326.
- Barton, N. H., & Hewitt, G. M. (1985). Analysis of hybrid zones. Annual Review of Ecology and Systematics, 16, 113–148. https://doi.org/10.1146/annurev.es.16.110185.000553
- Barton, N. H., & Hewitt, G. M. (1989). Adaptation, speciation and hybrid zones. *Nature*, 341, 497–503. https://doi.org/10.1038/341497a0
- Bates, D., Mächler, M., Bolker, B., & Walker, S. (2015). Fitting linear mixedeffects models using Ime4. *Journal of Statistical Software*, 67, 1–48.
- Beamish, R. J., & Northcote, T. G. (1989). Extinction of a population of anadromous parasitic lamprey, Lampetra tridentata, upstream of an impassable dam. Canadian Journal of Fisheries and Aquatic Sciences, 46, 420-425. https://doi.org/10.1139/f89-056
- Beier, P., & Noss, R. F. (1998). Do habitat corridors provide connectivity? Conservation Biology, 12, 1241–1252.
- Beninde, J., Veith, M., & Hochkirch, A. (2015). Biodiversity in cities needs space: A meta-analysis of factors determining intra-urban biodiversity variation. *Ecology Letters*, 18, 581–592. https://doi.org/10.1111/ele.12427

- Berdahl, A., Westley, P. A. H., Levin, S. A., Couzin, I. D., & Quinn, T. P. (2016). A collective navigation hypothesis for homeward migration in anadromous salmonids. *Fish and Fisheries*, 17, 525–542. https://doi. org/10.1111/faf.12084
- Berg, O. K. (1985). The formation of non-anadromous populations of Atlantic salmon, *Salmon salar* L., in Europe. *Journal of Fish Biology*, 27, 805–815. https://doi.org/10.1111/j.1095-8649.1985. tb03222.x
- Bernhardt, E. S., Palmer, M. A., Allan, J. D., Alexander, G., Barnas, K., Brooks, S., ... Sudduth, E. (2005). Synthesizing U.S. river restoration efforts. *Science*, 308, 636–637. https://doi.org/10.1126/science.1109769
- Bernhardt, E.S., Sudduth, E.B., Palmer, M.A., Allan, J.D., Meyer, J.L., Alexander, G., ... Pagano, L. (2007). Restoring rivers one reach at a time: Results from a survey of U.S. river restoration practitioners. *Restoration Ecology*, 15, 482–493. https://doi.org/10.1111/j.1526-100X.2007.00244.x
- Bicudo, T., Anciães, M., Benchimol, M., Peres, C. A., & Simões, P. I. (2016). Insularization effects on acoustic signals of 2 suboscine Amazonian birds. *Behavioral Ecology*, 27, 1480–1490. https://doi.org/10.1093/beheco/arw070
- Campbell, J. S. (1977). Spawning characteristics of brown trout and sea trout *Salmo trutta* L. in Kirk Burn, River Tweed, Scotland. *Journal of Fish Biology*, 11, 217–229. https://doi.org/10.1111/j.1095-8649.1977. tb04115.x
- Closs, G. P., Hicks, A. S., & Jellyman, P. G. (2013). Life histories of closely related amphidromous and non-migratory fish species: A trade-off between egg size and fecundity. Freshwater Biology, 58, 1162–1177. https://doi.org/10.1111/fwb.12116
- Coyne, J. A. (1992). Genetics and speciation. *Nature*, 355, 511-515. https://doi.org/10.1038/355511a0
- Coyne, J. A., & Orr, H. A. (2004). Speciation. Sunderland, MA: Sinauer Associates.
- Cyr, H., & Cyr, I. (2003). Temporal scaling of temperature variability from land to oceans. *Evolutionary Ecology Research*, *5*, 1183–1197.
- Edsall, T. A. (1970). The effect of temperature on the rate of development and survival of alewife eggs and larvae. *Transactions of the American Fisheries Society*, *99*, 376–380. https://doi.org/10.1577/1548-8659(1970)99<376:TEOTOT>2.0.CO;2
- Ellis, D., & Vokoun, J. C. (2009). Earlier spring warming of coastal streams and implications for alewife migration timing. North American Journal of Fisheries Management, 29, 1584–1589. https://doi.org/10.1577/ M08-181.1
- Fraser, D. J., Debes, P. V., Bernatchez, L., & Hutchings, J. A. (2014). Population size, habitat fragmentation, and the nature of adaptive variation in a stream fish. Proceedings of the Royal Society of London. Series B, Biological Sciences, 281, 20140370. https://doi.org/10.1098/ rspb.2014.0370
- Friedland, K. D. (1998). Ocean climate influences on critical Atlantic salmon (*Salmo salar*) life history events. *Canadian Journal of Fisheries and Aquatic Sciences*, 55, 119–130. https://doi.org/10.1139/d98-003
- Gao, X., Lin, P., Li, M., Duan, Z., & Liu, H. (2016). Impact of the Three Gorges Dam on the spawning stock and natural reproduction of Chinese sturgeon in the Changjiang River, China. Chinese Journal of Oceanology and Limnology, 34, 894–901. https://doi.org/10.1098/rspb.2014.0370
- Gardner, M. L. G. (1976). A review of factors which may influence the sea-age and maturation of Atlantic salmon *Salmo salar* L. *Journal of Fish Biology*, *9*, 289–327. https://doi.org/10.1111/j.1095-8649.1976. tb04680.x
- Glover, K. A., Solberg, M. F., McGinnity, P., Hindar, K., Verspoor, E., Coulson, M. W., ... Svåsand, T. (2017). Half a century of genetic interaction between farmed and wild Atlantic salmon: Status of knowledge and unanswered questions. Fish and Fisheries, 18, 890–927. https://doi.org/10.1111/faf.12214
- Goniea, T. M., Keefer, M. L., Bjornn, T. C., Peery, C. A., Bennett, D. H., & Stuehrenberg, L. C. (2006). Behavioral thermoregulation and slowed

- migration by adult fall chinook salmon in response to high Columbia River water temperatures. *Transactions of the American Fisheries Society*, 135, 408–419. https://doi.org/10.1577/T04-113.1
- Gosset, C., Rives, J., & Labonne, J. (2006). Effect of habitat fragmentation on spawning migration of brown trout (*Salmo trutta L.*). *Ecology of Freshwater Fish*, 15, 247–254. https://doi.org/10.1111/j.1600-0633.2006.00144.x
- Grant, G. (2001). Dam removal: Panacea or pandora for rivers? Hydrological Processes, 15, 1531–1532. https://doi.org/10.1002/ (ISSN)1099-1085
- Gross, R. W. (1959). A study of the alewife Alosa pseudoharengus (Wilson) in some New Jersey lakes, with special reference to Lake Hopatcong. Master's Thesis, Rutgers State University, New Brunswick, NJ.
- Gulseth, O. A., & Nilssen, K. J. (2001). Life-history traits of charr, Salvelinus alpinus, from a high arctic watercourse on Svalbard. Arctic, 54, 1-11.
- Hall, C. J., Jordaan, A., & Frisk, M. G. (2012). Centuries of anadromous forage fish loss: Consequences for ecosystem connectivity and productivity. *BioScience*, 62, 723–731. https://doi.org/10.1525/bio.2012.62.8.5
- Hansen, L. P., Jonsson, N., & Jonsson, B. (1993). Oceanic migration in homing Atlantic salmon. Animal Behaviour, 45, 927-941. https://doi. org/10.1006/anbe.1993.1112
- Harnish, R. A., Sharma, R., McMichael, G. A., Langshaw, R. B., & Pearsons, T. N. (2014). Effect of hydroelectric dam operations on the freshwater productivity of a Columbia River fall Chinook salmon population. Canadian Journal of Fisheries and Aquatic Sciences, 71, 602–615. https://doi.org/10.1139/cjfas-2013-0276
- Hart, D. D., Johnson, T. E., Bushaw-Newton, K. L., Horwitz, R. J., Bednarek, A. T., Charles, D. F., ... Velinsky, D. J. (2002). Dam removal: Challenges and opportunities for ecological research and river restoration. *BioScience*, 52, 669-682. https://doi. org/10.1641/0006-3568(2002)052[0669:DRCAOF]2.0.CO;2
- Hasselman, D. J., Argo, E. E., Mcbride, M. C., Bentzen, P., Schultz, T. F., Perez-Umphrey, A. A., & Palkovacs, E. P. (2014). Human disturbance causes the formation of a hybrid swarm between two naturally sympatric fish species. *Molecular Ecology*, 23, 1137–1152. https://doi. org/10.1111/mec.12674
- Hasselman, D. J., & Limburg, K. E. (2012). Alosine restoration in the 21st century: Challenging the status quo. *Marine and Coastal Fisheries*, 4, 174–187. https://doi.org/10.1080/19425120.2012.675968
- Hewitt, G. M. (1988). Hybrid zones Natural laboratories for evolutionary studies. Trends in Ecology & Evolution, 3, 158–167. https://doi.org/10.1016/0169-5347(88)90033-X
- Hobbs, J. A., Munday, P. L., & Jones, G. P. (2004). Social induction of maturation and sex determination in a coral reef fish. Proceedings of the Royal Society of London. Series B, Biological Sciences, 271, 2109–2114. https://doi.org/10.1098/rspb.2004.2845
- Hodgson, S., & Quinn, T. P. (2002). The timing of adult sockeye salmon migration into fresh water: Adaptations by populations to prevailing thermal regimes. *Canadian Journal of Zoology*, 80, 542–555. https:// doi.org/10.1139/z02-030
- Hougaard, P. (2000). Analysis of multivariate survival data (1st ed.). New York, NY: Springer-Verlag. https://doi.org/10.1007/978-1-4612-1304-8
- Hutchings, J. A., & Myers, R. A. (1985). Mating between anadromous and nonanadromous Atlantic salmon, Salmo salar. Canadian Journal of Zoology, 63, 2219–2221. https://doi.org/10.1139/z85-327
- Jones, F. C., Brown, C., Pemberton, J. M., & Braithwaite, V. A. (2006). Reproductive isolation in a threespine stickleback hybrid zone. Journal of Evolutionary Biology, 19, 1531–1544. https://doi. org/10.1111/j.1420-9101.2006.01122.x
- Jones, A. W., Palkovacs, E. P., & Post, D. M. (2013). Recent parallel divergence in body shape and diet source of alewife life history forms. Evolutionary Ecology, 27, 1175–1187. https://doi.org/10.1007/s10682-013-9650-2

- Karve, A. D., von Hippel, F. A., & Bell, M. A. (2008). Isolation between sympatric anadromous and resident threespine stickleback species in Mud Lake, Alaska. Environmental Biology of Fishes, 81, 287–296.
- Kissil, G. W. (1974). Spawning of the anadromous alewife, Alosa pseudoharengus, in Bride Lake, Connecticut. Transactions of the American Fisheries Society, 103, 312–317. https://doi.org/10.1577/1548-8659 (1974)103<312:SOTAAA>2.0.CO;2
- Kovach, R. P., Muhlfeld, C. C., Boyer, M. C., Lowe, W. H., Allendorf, F. W., & Luikart, G. (2015). Dispersal and selection mediate hybridization between a native and invasive species. *Proceedings of the Royal Society B-Biological Sciences*, 282, 20142454. https://doi.org/10.1098/rspb.2014.2454
- Lackey, R. T. (1970). Observations on newly introduced landlocked alewives in Maine. New York Fish and Game Journal, 17, 110-116.
- Lake, T. R. T., Ravana, K. R., & Saunders, R. (2012). Evaluating changes in diadromous species distributions and habitat accessibility following the Penobscot River restoration project. *Marine and Coastal Fisheries*, 4, 284–293. https://doi.org/10.1080/19425120.2012.67 5971
- Lenhart, C. F. (2003). A preliminary review of NOAA's community-based dam removal and fish passage projects. *Coastal Management*, 31, 79–98. https://doi.org/10.1080/08920750390168318
- Levine, J. M., & HilleRisLambers, J. (2009). The importance of niches for the maintenance of species diversity. *Nature*, 461, 254–257. https://doi.org/10.1038/nature08251
- Liermann, C. R., Nilsson, C., Robertson, J., & Ng, R. Y. (2012). Implications of dam obstruction for global freshwater fish diversity. *BioScience*, 62, 539–548. https://doi.org/10.1525/bio.2012.62.6.5
- Limburg, K. E., & Waldman, J. R. (2009). Dramatic declines in north Atlantic diadromous fishes. *BioScience*, 59, 955–965. https://doi. org/10.1525/bio.2009.59.11.7
- Locke, A., Hanson, J. M., Klassen, G. J., Richardson, S. M., & Aubé, C. I. (2003). The damming of the Petitcodiac River: Species, populations, and habitats lost. *Northeastern Naturalist*, 10, 39–54. https://doi.org/10.1656/1092-6194(2003)010[0039:TDOTPR]2.0. CO;2
- Lyons, J., Rypel, A. L., Rasmussen, P. W., Burzynski, T. E., Eggold, B. T., Myers, J. T., ... McIntyre, P. B. (2015). Trends in the reproductive phenology of two Great Lakes fishes. *Transactions of the American Fisheries Society*, 144, 1263–1274. https://doi.org/10.1080/00028487.2015.1082502
- Mayfield, M. M., & Levine, J. M. (2010). Opposing effects of competitive exclusion on the phylogenetic structure of communities. *Ecology Letters*, 13, 1085–1093. https://doi.org/10.1111/j.1461-0248.2010.01509.x
- McDowall, R. M. (1988). Diadromy in fishes: Migrations between freshwater and marine environments. London and Sydney: Croom Helm.
- Neves, R. J. (1981). Offshore distribution of alewife, *Alosa pseudoharengus*, and blueback herring, *Alosa aestivalis*, along the Atlantic coast. *Fishery Bulletin*, *79*, 473–485.
- Nigro, A. A., & Ney, J. J. (1982). Reproduction and early-life accommodations of landlocked alewives to a southern range extension. Transactions of the American Fisheries Society, 111, 559-569. https://doi.org/10.1577/1548-8659(1982)111<559:RAEAOL>2.0.CO;2
- NOAA Tides and Currents (2018). Water temperature at 8461490, New London, Thames River, CT. Retrieved from https://tidesandcurrents.noaa.gov/physocean.html?id=8461490
- Odell, T. T. (1934). The life history and ecological relationships of the alewife (*Pomolobus pseudoharengus* (Wilson) in Seneca Lake, New York. Transactions of the American Fisheries Society, 64, 118–126. https://doi.org/10.1577/1548-8659(1934)64[118:TLHAER]2.0.CO;2
- Palkovacs, E. P., Dion, K. B., Post, D. M., & Caccone, A. (2008). Independent evolutionary origins of landlocked alewife populations and rapid parallel evolution of phenotypic traits. *Molecular Ecology*, 17, 582–597.

- Palkovacs, E. P., Mandeville, E. G., & Post, D. M. (2014). Contemporary trait change in a classic ecological experiment: Rapid decrease in alewife gill-raker spacing following introduction to an inland lake. Freshwater Biology, 59, 1897–1901. https://doi.org/10.1111/ fwb.12392
- Palkovacs, E. P., & Post, D. M. (2009). Experimental evidence that phenotypic divergence in predators drives community divergence in prey. *Ecology*, 90, 300–305. https://doi.org/10.1890/08-1673.1
- Pearse, D. E., Hayes, S. A., Bond, M. H., Hanson, C. V., Anderson, E. C., Macfarlane, R. B., & Garza, J. C. (2009). Over the falls? Rapid evolution of ecotypic differentiation in steelhead/rainbow trout (Oncorhynchus mykiss). Journal of Heredity, 100, 515–525. https://doi.org/10.1093/jhered/esp040
- Perry, W. L., Feder, J. L., Dwyer, G., & Lodge, D. M. (2001). Hybrid zone dynamics and species replacement between *Orconectes* crayfishes in a northern Wisconsin Lake. *Evolution*, *55*, 1153–1166. https://doi.org/10.1111/j.0014-3820.2001.tb00635.x
- Pohl, M. M. (2002). Bringing down our dams: Trends in American dam removal rationales. *Journal of the American Water Resources Association*, 38, 1511–1519. https://doi.org/10.1111/j.1752-1688.2002. tb04361.x
- Post, D. M., Palkovacs, E. P., Schielke, E. G., & Dodson, S. I. (2008). Intraspecific variation in a predator affects community structure and cascading trophic interactions. *Ecology*, 89, 2019–2032. https://doi.org/10.1890/07-1216.1
- Potts, W. M., Henriques, R., Santos, C. V., Munnik, K., Ansorge, I., Dufois, F., ... Shaw, P. W. (2014). Ocean warming, a rapid distributional shift, and the hybridization of a coastal fish species. *Global Change Biology*, 20, 2765–2777. https://doi.org/10.1111/gcb.12612
- Pritchard, A. L. (1929). The alewife (Pomolobus pseudoharengus) in Lake Ontario (Vol. 38). Toronto, ON: Ontario Fisheries Research Lab.
- Quinn, T. P., & Adams, D. J. (1996). Environmental changes affecting the migratory timing of American shad and sockeye salmon. *Ecology*, 77, 1151–1162. https://doi.org/10.2307/2265584
- Quinn, T. P., Hodgson, S., & Peven, C. (1997). Temperature, flow, and the migration of adult sockeye salmon (*Oncorhynchus nerka*) in the Columbia River. *Canadian Journal of Fisheries and Aquatic Sciences*, 54, 1349–1360. https://doi.org/10.1139/f97-038
- Quinn, T. P., McGinnity, P., & Reed, T. E. (2016). The paradox of "premature migration" by adult anadromous salmonid fishes: Patterns and hypotheses. Canadian Journal of Fisheries and Aquatic Sciences, 73, 1015–1030.
- R Core Team (2017). R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. Retrieved from https://www.R-project.org/
- Rand, P. S., Hinch, S. G., Morrison, J., Foreman, M. G. G., MacNutt, M. J., Macdonald, J. S., ... Higgs, D. A. (2006). Effects of river discharge, temperature, and future climates on energetics and mortality of adult migrating Fraser River sockeye salmon. *Transactions of the American Fisheries Society*, 135, 655–667. https://doi.org/10.1577/T05-023.1
- Resasco, J., Bruna, E. M., Haddad, N. M., Banks-Leite, C., & Margules, C. R. (2017). The contribution of theory and experiments to conservation in fragmented landscapes. *Ecography*, 40, 109–118.
- Riva-Rossi, C., Pascual, M. A., Babaluk, J. A., García-Asorey, M., & Halden, N. M. (2007). Intra-population variation in anadromy and reproductive life span in rainbow trout introduced in the Santa Cruz River, Argentina. *Journal of Fish Biology*, 70, 1780–1797. https://doi.org/10.1111/j.1095-8649.2007.01449.x
- Rivers, A. (2017). American rivers dam removal database. https://doi.org/10.6084/m9.figshare.5234068.v2
- Rolls, R. J., Stewart-Koster, B., Ellison, T., Faggotter, S., & Roberts, D. T. (2014). Multiple factors determine the effect of anthropogenic barriers to connectivity on riverine fish. *Biodiversity and Conservation*, 23, 2201–2220. https://doi.org/10.1007/s10531-014-0715-5

- Rosset, J., Roy, A. H., Gahagan, B. I., Whiteley, A. R., Armstrong, M. P., Sheppard, J. J., & Jordaan, A. (2017). Temporal patterns of migration and spawning of river herring in coastal Massachusetts. *Transactions* of the American Fisheries Society, 146, 1101–1114. https://doi.org/10. 1080/00028487.2017.1341851
- Rothschild, B. J. (1966). Observations on the alewife (Alosa pseudoharengus) (Wilson) in Cayuga Lake. New York Fish and Game Journal, 13, 188–195.
- Santos, A. B. I., & Araújo, F. G. (2015). Evidence of morphological differences between Astyanax bimaculatus (Actinopterygii: Characidae) from reaches above and below dams on a tropical river. Environmental Biology of Fishes, 98, 183–191. https://doi.org/10.1007/s10641-014-0248-5
- Schielke, E. G., Palkovacs, E. P., & Post, D. M. (2011). Eco-evolutionary feedbacks drive niche differentiation in the alewife. *Biological Theory*, 6, 211–219. https://doi.org/10.1007/s13752-012-0031-9
- Seehausen, O., van Alphen, J. J. M., & Witte, F. (1997). Cichlid fish diversity threatened by eutrophication that curbs sexual selection. Science, 277, 1808–1811. https://doi.org/10.1126/science.277.5333 .1808
- Sun, J. (2006). The statistical analysis of interval-censored failure time data (1st ed.). New York, NY: Springer-Verlag.
- Tulp, I., Keller, M., Navez, J., Winter, H. V., de Graaf, M., & Baeyens, W. (2013). Connectivity between migrating and landlocked populations of a diadromous fish species investigated using otolith microchemistry. PLoS One, 8, e69796. https://doi.org/10.1371/journal.pone.0069796
- Twining, C. W., & Post, D. M. (2013). Cladoceran remains reveal presence of a keystone size-selective planktivore. *Journal of Paleolimnology*, 49, 253–266. https://doi.org/10.1007/s10933-012-9672-8
- U.S. Fish & Wildlife Service (2012). National fish passage program: creating aquatic possibilities. U.S. Fish & Wildlife Service. Retrieved from https://www.fws.gov/alaska/fisheries/restoration/pdf/2012_national_fish_passage_program.pdf
- Velotta, J. P., McCormick, S. D., Jones, A. W., & Schultz, E. T. (2018). Reduced swimming performance repeatedly evolves on loss of migration in landlocked populations of alewife. *Physiological* and Biochemical Zoology, 91, 814–825. https://doi.org/10.1086/ 696877
- Velotta, J. P., McCormick, S. D., O'Neill, R. J., & Schultz, E. T. (2014). Relaxed selection causes microevolution of seawater osmoregulation and gene expression in landlocked alewives. *Oecologia*, 175, 1081–1092. https://doi.org/10.1007/s00442-014-2961-3
- Velotta, J. P., McCormick, S. D., & Schultz, E. T. (2015). Trade-offs in osmoregulation and parallel shifts in molecular function follow ecological transitions to freshwater in the alewife. *Evolution*, 69, 2676–2688. https://doi.org/10.1111/evo.12774
- Vörösmarty, C. J., McIntyre, P. B., Gessner, M. O., Dudgeon, D., Prusevich, A., Green, P., ... Davies, P. M. (2010). Global threats to human water security and river biodiversity. *Nature*, 467, 555–561. https://doi. org/10.1038/nature09440
- Zaccara, S., Antognazza, C. M., Buonerba, L., Britton, R., & Crosa, G. (2014). Human-mediated contact zone between endemic and invasive *Barbus* species (Osteichthyes: *Cyprinidae*) in a regulated low-land river: Genetic inferences and conservation implications. *Italian Journal of Zoology*, 81, 571–583. https://doi.org/10.1080/11250003.2014.944225
- Zastavniouk, C., Weir, L. K., & Fraser, D. J. (2017). The evolutionary consequences of habitat fragmentation: Body morphology and coloration differentiation among brook trout populations of varying size. *Ecology and Evolution*, 7, 6850–6862. https://doi.org/10.1002/ece3.3229
- Zhou, J., Zhao, Y., Song, L., Bi, S., & Zhang, H. (2014). Assessing the effect of the Three Gorges reservoir impoundment on spawning habitat suitability of Chinese sturgeon (*Acipenser sinensis*) in Yangtze River,

Evolutionary Applications — WILEY 13

China. Ecological Informatics, 20, 33–46. https://doi.org/10.1016/j.ecoinf.2014.01.008

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

How to cite this article: Littrell KA, Ellis D, Gephard SR, et al. Evaluating the potential for prezygotic isolation and hybridization between landlocked and anadromous alewife (*Alosa pseudoharengus*) following secondary contact. *Evol Appl.* 2018;00:1–13. https://doi.org/10.1111/eva.12645